



# Role of Biotechnological Interventions in Improving the Traits of Flowering Ornamentals

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# **ABSTRACT**

Improving the floricultural traits in ornamentals is of great value in floriculture and landscaping. Conventional methods have been and are being still used for improving the floricultural traits like vase life or floral longevity, ethylene sensitivity and visible floral morphological features, etc. in ornamentals by postharvest application of ethylene antagonists, plant growth regulators, sugar sources, protein synthesis inhibitors and antimicrobial compounds, etc. However, biotechnology offers a promising approach to improving the desirable attributes in flowering ornamentals like flower color, fragrance, longevity, ethylene insensitivity, disease resistance, etc. In the present review, the biotechnological interventions in enhancing the flower fragrance, flower color and flower longevity/vase life have been discussed and a number of suitable examples related to the improvement of these traits in various ornamentals have been provided. The mini-review intends to present a comprehensive update of the available literature regarding the improvement of these traits in flowering ornamentals involving the non-conventional biotechnological approach.

# **KEYWORDS**

Biotechnology, flower color, fragrance, longevity, transgenics, ornamentals, floriculture

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# **INTRODUCTION**

Floriculture or flower farming, one of the important horticultural disciplines, primarily focuses on various parameters related to ornamentals that include their propagation, cultivation, harvesting, as well as postharvest processing. Efforts have been made and are being made to improve the floral attributes so as to make them more market-oriented. The research leading to the improvement in the attributes of these ornamentals is being conducted at different levels-preharvest level, harvest and postharvest level. The preharvest level research parameters include the quality of the plant material (seed, bulb, rhizome, etc.), appropriate growing conditions (soil requirements, fertilizer application, water requirements, light conditions and temperature etc.) and protection from diseases and pests. Appropriate time of harvest, the right method and the right stage of harvest are the primary factors influencing the harvest stage. The postharvest level factors that influence the attributes of ornamentals include proper transportation and storage, use of chemical preservatives and growth regulators as pulse or spray treatments, maintenance of sugar supply, use of antimicrobial agents in vase solutions, use of ethylene antagonists and optimum ranges of temperature and relative humidity, etc. A number of floral



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attributes like size, longevity and quality have been improved to a great extent by research at pre-harvest as well as postharvest level. Moreover, the intervention of biotechnology or genetic engineering has offered a promising approach for developing new ornamentals with improved floral attributes as well as producing ornamentals with novel traits. Altered flower colour, enhanced fragrance, disease resistance, ethylene in-sensitivity (in ethylene-sensitive flowers) and early blooming are some of the important attributes that have been targeted using biotechnological interventions<sup>1-4</sup>. The present review intends to provide comprehensive information about the role of biotechnology in improving and modifying the attributes of ornamentals with the purpose to enhance their commercial value.

**Flower fragrance:** The flower fragrance has been attributed to the biosynthesis of many volatile compounds like terpenoids, benzenoids, phenylpropanoid and other Sulphur or Nitrogen-containing compounds<sup>5</sup>. Floral fragrance or scent production in ornamentals offers many advantages like a signal for pollinators, attraction for predators of herbivores and repulsion of herbivores, etc<sup>5-7</sup>. A number of ornamentals have been genetically modified to enhance scent production. Some of the notable examples were listed in Table 1.

From Table 1, it is clear that scent production has been enhanced in the ornamentals by using two approaches i.e. either by inducing the production of volatile compounds like Benzyl acetate and S-linalool or by regulating the expression of the *F3'H* gene (downregulation in Carnation) or *PAP1* gene (Overexpression in Rose). Here, it deserves mention that the *AtPAP1* gene is responsible for flower color modification as well as for enhancing the fragrance.

**Flower color:** Another important attribute of ornamentals is the floral display in the form of various colors which has been attributed to the occurrence of different plant pigments in the floral organs. These pigments include anthocyanins, chlorophylls, carotenoids, betalains, etc. These pigments are known to play important roles like attraction to pollinators, light capturing for photosynthesis and protection from UV radiation besides determining the ornamental value of the plants<sup>12</sup>. Many ornamentals have been genetically engineered to produce flowers with novel colors or enhanced colors. Some of the notable examples were listed in Table 2.

A number of flowers with novel or enhanced flower colors have been obtained using the biotechnological approach (Table 2) wherein the genes like *DFR gene*, *F3'H* genes, *F3'5'H* genes and *PAP1* gene have been introduced to host ornamentals and made to express and produce the novel phenotype. Moreover, the silencing or down regulation of genes like the CHS gene (Chalcone synthase), ANS gene, *F3'5'H* genes and the overexpression of the *PAP1* gene and CrtW gene have been found to enhance or modify the flower color in many ornamentals.

Vase life/shelf life: Another important parameter that determines the postharvest quality of ornamentals is their vase life or shelf life. Flower longevity or vase life determines how long a flower or a spike or scape remains in fresh condition after being detached from the mother plant, particularly in the case of cut flowers. A number of preharvest and postharvest treatments are known to enhance the vase life or shelf life of ornaments by conventional use of Plant Growth Regulators (PGRs) and vase solutions containing

Table 1: Genetic modification in ornamentals for flower fragrance

Plant	Gene	Source	References
Carnation and lisianthus	LIS gene that codes S-linalool synthase Clarkia brewer		Zaccai et al.8
Carnation	Downregulation of the F3H gene	Carnation	Zuker et al.9
Lisianthus	Benzyl alcohol acetyl transferase (BEAT) for the production of benzyl acetate	Clarkia breweri	Aranovich et al. <sup>10</sup>
Rosa hybrida	Overexpression of the AtPAP1 gene	Arabidopsis thaliana	Zvi et al. <sup>11</sup>

Table 2: Genetic modification in ornamentals for flower color

Plant	Gene	Source	Altered flower color	Reference number
Petunia (deficient in	DFR gene	Maize	Orange	Meyer et al.13 and Meyer et al.1
F3′5′H and F3′H)				Forkmann et al.15
Chrysanthemum	Expressing sense or	-	White or light pink	Courtney-Gutterson et al.16
(Pink)	antisense copies of			
	the Chs gene encoding			
	chalcone synthase			
Carnation (White)	F3'5'H and DFR gene	Petunia	Blue	Lou et al. <sup>17</sup>
			(FLORIGEN)	
			E®Moondust™)	
Carnation (white)	F3'5'H and DFR gene	Viola and Petunia,	Dark violet	Mol et al. <sup>18</sup>
		respectively	FLORIGENE®	
			Moonshadow™	
Arabidopsis and	Overexpression of	Arabidopsis AtMYB75,	Flowers with enhanced	Borevitz <i>et al</i> . <sup>19</sup>
Tobacco	PAP1	(PAP1 transcription	pigmentation	
		factor)		
Carnation	Antisense gene	Carnation	White	Zuker et al. <sup>9</sup>
	construct for F3'H gene			
Petunia	Down regulation of	Rosa	Red	Tsuda et al. <sup>20</sup>
	F3'H gene (petunia)			
	and over expression			
	of rose <i>DFR</i> gene			
Rose	F3'5'H gene	Viola	Blue rose	Katsumoto et al. <sup>21</sup>
Torenia	Anthocyanin biosynthesis	Torenia	White	Nakamura et al. <sup>22</sup>
	genes (by employing RNAi)			
Torenia (Blue)	Down regulation of	Pelargonium	Pink	Nakatsuka et al. <sup>23</sup>
	F3'5'H and F3'H genes and			
	over expression of			
	Pelargonium <i>DFR</i> gene			22
Tobacco	Down regulation of	Gerbera	Red	Nakatsuka et al. <sup>23</sup>
	F3'H gene and			
	flavonol synthase gene			
	and overexpression of			
	gerbera <i>DFR</i> gene			24
Lotus japonica	Overexpression	Lotus japonica	Deep yellow to	Suzuki <i>et al</i> . <sup>24</sup>
	of CrtW gene		orange	25
Gentiana	Suppression of the	Gentiana	Magenta	Nakatsuka et al. <sup>25</sup>
	F3'5'H gene			
Gentian	Silencing of CHS	Gentian	White and pale blue,	Nakatsuka et al.25 and
	and ANS genes		respectively	Yoshida <i>et al</i> . <sup>26</sup>
Gentian	Downregulating the	Gentian	Pink	Nakatsuka et al. <sup>25</sup>
	F3'5'H gene and the			
	5,3'-AT gene (encoding			
	anthocyanin)			
Chrysanthemum	Expressing F3'5'H gene	-	Blue	Nakatsuka et al. <sup>27</sup>
	under rose CHS promoter			
Chrysanthemum	Suppression of CmCCD4a	Chrysanthemum	Yellow	Ohmiya et al. <sup>28</sup> and
	(Carotenoid cleavage		(Yellow Jimba)	Ohmiya et al. <sup>29</sup>
	dioxygenase) gene			
Lilium	PhF3'5'H gene	Phalaenopsis	Purple	Azadi et al. <sup>30</sup>
Cyclamen persicum	Suppression of the	Cyclamen	Red/Pink	Boase et al. <sup>31</sup>
(Purple)	F3'5'H gene			
Chrysanthemum	Suppression of	Chrysanthemum	Bright red	He et al. <sup>32</sup>
-	CmF3′H gene	-	-	

sugars, sugar alcohols, biocides, ethylene antagonists and other chemicals<sup>33,34</sup>. However, genetic engineering has also been employed to produce ornamentals with enhanced shelf life/vase life. Some of the notable examples were listed in Table 3.

Table 3: Genetic modification in ornamentals for enhanced vase life

Plant	Gene	Source	References
Carnation	Silencing the ACO gene	-	Savin et al. <sup>35</sup>
Carnation	etr1-1 gene Arabidopsis		Bovy et al. <sup>36</sup>
Petunia	Transformation of etr1-1 gene	Arabidopsis	Gubrium et al. <sup>37</sup>
Chrysanthemum			Ma et al. <sup>38</sup>
Carnation	ACO gene and ACS gene	Carnation and apple	Inokuma et al. <sup>39</sup>
Oncidium	Mutating ethylene receptor gene	-	Raffeiner et al.40
Odontoglossum	Mutating ethylene receptor gene	-	Raffeiner et al. <sup>40</sup>

Another feature added to the cap of ornamental biotechnology is the development of fluorescent flowers in Torenia by using a yellowish-green fluorescent gene-*CpYGFP* from the marine plankton *Chiridius poppei*<sup>41</sup>. These fluorescent ornamentals serve the dual purpose-one being the enhancement of the ornamental value of the plant and the other being used for the analysis of fluorescent transgenic plants spatiotemporally in a non-destructive manner.

# **CONCLUSION**

In conclusion, biotechnology involving the genetic engineering approach has not only provided a significant contribution to modifying or improving the existing traits of flowering ornamentals but has led to the production of ornamentals with novel traits. These modifications in ornamentals have been proven to be a successful venture from scientific as well as commercial points of view. As far as the future perspective of ornamental horticulture is concerned, biotechnology has an important role to play which involves the extension of the existing biotechnological strategies/approaches to other related ornamentals and to promote research in more areas related to flowering ornamentals.

# SIGNIFICANCE STATEMENT

Biotechnology offers a promising approach to improve the traits of the ornamentals when compared to traditional methods of ornamental plant improvement. Using the biotechnological approach, many milestones have been achieved while dealing with ornamentals like enhancement of vase life or flower longevity, enhancing the floral fragrance/scent, modifying the flower color or producing flowers with novel colors and reducing sensitivity to ethylene, etc. The purpose of the mini-review was to present an update on the improvement of some selected traits in flowering ornamentals using the biotechnological approach. Many examples of genetic modifications in ornamentals have been provided in the review so as to comprehend the importance of biotechnology in ornamental horticulture.

### **REFERENCES**

- 1. Chandler, S.F. and C. Sanchez, 2012. Genetic modification; the development of transgenic ornamental plant varieties. Plant Biotechnol. J., 10: 891-903.
- 2. Parisi, C., P. Tillie and E. Rodríguez-Cerezo, 2016. The global pipeline of GM crops out to 2020. Nat. Biotechnol., 34: 31-36.
- 3. Noman, A., R. Bashir, M. Aqeel, S. Anwer and W. Iftikhar *et al.*, 2016. Success of transgenic cotton (*Gossypium hirsutum* L.): Fiction or reality? Cogent Food Agric., Vol. 2. 10.1080/23311932.2016.1207844.
- 4. Noman, A., M. Aqeel, J. Deng, N. Khalid, T. Sanaullah and H. Shuilin, 2017. Biotechnological advancements for improving floral attributes in ornamental plants. Front. Plant Sci., Vol. 8. 10.3389/fpls.2017.00530.
- 5. Knudsen, J.T., L. Tollsten and L.G. Bergström, 1993. Floral scents-a checklist of volatile compounds isolated by head-space techniques. Phytochemistry, 33: 253-280.
- 6. Gershenzon, J. and R. Croteau, 1991. Terpenoids. In: Herbivores: Their Interactions with Secondary Plant Metabolites, Rosenthal, G.A. and M.R. Berenbaum (Eds.), Academic Press, United States, ISBN: 978-0-12-597183-6, pp: 165-219.

- 7. Pare, P.W. and J.H. Tumlinson, 1997. De novo biosynthesis of volatiles induced by insect herbivory in cotton plants. Plant Physiol., 114: 1161-1167.
- 8. Zaccai, M., E. Lewinsohn and E. Pichersky, 2001. Modifying *Lisianthus* traits by genetic engineering. Acta Hortic., 552: 137-142.
- 9. Zuker, A., T. Tzfira, H. Ben-Meir, M. Ovadis and E. Shklarman *et al.*, 2002. Modification of flower color and fragrance by antisense suppression of the flavanone 3-hydroxylase gene. Mol. Breed., 9: 33-41.
- 10. Aranovich, D., E. Lewinsohn and M. Zaccai, 2007. Post-harvest enhancement of aroma in transgenic lisianthus (*Eustoma grandiflorum*) using the *Clarkia breweri* benzyl alcohol acetyltransferase (BEAT) gene. Postharvest Biol. Technol., 43: 255-260.
- 11. Zvi, M.M.B., E. Shklarman, T. Masci, H. Kalev and T. Debener *et al.*, 2012. *PAP1* transcription factor enhances production of phenylpropanoid and terpenoid scent compounds in rose flowers. New Phytol., 195: 335-345.
- 12. Davies, K.M., K.E. Schwinn, S.C. Deroles, D.G. Manson, D.H. Lewis, S.J. Bloor and J.M. Bradley, 2003. Enhancing anthocyanin production by altering competition for substrate between flavonol synthase and dihydroflavonol 4-reductase. Euphytica, 131: 259-268.
- 13. Meyer, P., I. Heidmann, G. Forkmann and H. Saedler, 1987. A new petunia flower colour generated by transformation of a mutant with a maize gene. Nature, 330: 677-678.
- 14. Meyer, P., F. Linn, I. Heidmann, H. Meyer, I. Niedenhof and H. Saedler, 1992. Endogenous and environmental factors influence 35S promoter methylation of a maize A1 gene construct in transgenic petunia and its colour phenotype. Mole. Gen. Genet. MGG, 231: 345-352.
- 15. Forkmann, G. and B. Ruhnau, 1987. Distinct substrate specificity of dihydroflavonol 4-reductase from flowers of petunia hybrida. Z. für Naturforsch. C, 42: 1146-1148.
- 16. Courtney-Gutterson, N., C. Napoli, C. Lemieux, A. Morgan, E. Firoozabady and K.E.P. Robinson, 1994. Modification of flower color in florist's chrysanthemum: Production of a white-flowering variety through molecular genetics. Nat. Biotechnol., 12: 268-271.
- 17. Lou, Y., Q. Zhang, Q. Xu, X. Yu, W. Wang, R. Gai and F. Ming, 2023. PhCHS5 and PhF3'5'H genes over-expression in *Petunia (Petunia hybrida)* and phalaenopsis (*Phalaenopsis aphrodite*) regulate flower color and branch number. Plants, Vol. 12. 10.3390/plants12112204.
- 18. Mol, J., E. Cornish, J. Mason and R. Koes, 1999. Novel coloured flowers. Curr. Opin. Biotechnol., 10: 198-201.
- 19. Borevitz, J.O., Y. Xia, J. Blount, R.A. Dixon and C. Lamb, 2000. Activation tagging identifies a conserved MYB regulator of phenylpropanoid biosynthesis. Plant Cell, 12: 2383-2393.
- 20. Tsuda, S., Y. Fukui, N. Nakamura, Y. Katsumoto and K. Yonekura-Sakakibara *et al.*, 2004. Flower color modification of *Petunia hybrida* commercial varieties by metabolic engineering. Plant Biotechnol., 21: 377-386.
- 21. Katsumoto, Y., M. Fukuchi-Mizutani, Y. Fukui, F. Brugliera and T.A. Holton *et al.*, 2007. Engineering of the rose flavonoid biosynthetic pathway successfully generated blue-hued flowers accumulating delphinidin. Plant Cell Physiol., 48: 1589-1600.
- 22. Nakamura, N., M. Fukuchi-Mizutani, K. Miyazaki, K. Suzuki and Y. Tanaka, 2006. RNAi suppression of the anthocyanidin synthase gene in *Torenia hybrida* yields white flowers with higher frequency and better stability than antisense and sense suppression. Plant Biotechnol., 23: 13-17.
- 23. Nakatsuka, T., Y. Abe, Y. Kakizaki, S. Yamamura and M. Nishihara, 2007. Production of red-flowered plants by genetic engineering of multiple flavonoid biosynthetic genes. Plant Cell Rep., 26: 1951-1959.
- 24. Suzuki, S., M. Nishihara, T. Nakatsuka, N. Misawa, I. Ogiwara and S. Yamamura, 2007. Flower color alteration in *Lotus japonicus* by modification of the carotenoid biosynthetic pathway. Plant Cell Rep., 26: 951-959.
- 25. Nakatsuka, T., K.I. Mishiba, Y. Abe, A. Kubota, Y. Kakizaki, S. Yamamura and M. Nishihara, 2008. Flower color modification of gentian plants by RNAi-mediated gene silencing. Plant Biotechnol., 25: 61-68.

- 26. Yoshida, K., M. Mori and T. Kondo, 2009. Blue flower color development by anthocyanins: From chemical structure to cell physiology. Nat. Prod. Rep., 26: 884-915.
- 27. Nakatsuka, T., K.I. Mishiba, A. Kubota, Y. Abe and S. Yamamura *et al.*, 2010. Genetic engineering of novel flower colour by suppression of anthocyanin modification genes in gentian. J. Plant Physiol., 167: 231-237.
- 28. Ohmiya, A., S. Kishimoto, R. Aida, S. Yoshioka and K. Sumitomo, 2006. Carotenoid cleavage dioxygenase (CmCCD4a) contributes to white color formation in chrysanthemum petals. Plant Physiol., 142: 1193-1201.
- 29. Ohmiya, A., K. Sumitomo and R. Aida, 2009. "Yellow Jimba": Suppression of carotenoid cleavage dioxygenase (*CmCCD4a*) expression turns white chrysanthemum petals yellow. J. Japan. Soc. Hort. Sci., 78: 450-455.
- 30. Azadi, P., N.V. Otang, D.P. Chin, I. Nakamura and M. Fujisawa *et al.*, 2010. Metabolic engineering of *Lilium* × *formolongi* using multiple genes of the carotenoid biosynthesis pathway. Plant Biotechnol. Rep., 4: 269-280.
- 31. Boase, M.R., D.H. Lewis, K.M. Davies, G.B. Marshall, D. Patel, K.E. Schwinn and S.C. Deroles, 2010. Isolation and antisense suppression of *flavonoid 3'*, *5'-hydroxylase* modifies flower pigments and colour in cyclamen. BMC Plant Biol., Vol. 10. 10.1186/1471-2229-10-107.
- 32. He, H., H. Ke, H. Keting, X. Qiaoyan and D. Silan, 2013. Flower colour modification of chrysanthemum by suppression of *F3'H* and overexpression of the exogenous *Senecio cruentus F3'5'H* gene. PLoS ONE, Vol. 8. 10.1371/journal.pone.0074395.
- 33. Shahri, W., I. Tahir, S.T. Islam and M. Ahmad, 2010. Response of some ornamental flowers of family Ranunculaceae to sucrose feeding. Afr. J. Plant Sci., 4: 346-352.
- 34. Gul, F., I. Tahir and W. Shahri, 2015. Improvement in postharvest performance of cut scapes of *Nerine sarniensis* red by the application of growth regulators. Int. J. Postharvest Technol. Innovation, 4: 103-113.
- 35. Savin, K.W., S.C. Baudinette, M.W. Graham, M.Z. Michael and G.D. Nugent *et al.*, 1995. Antisense ACC oxidase RNA delays carnation petal senescence. HortScience, 30: 970-972.
- 36. Bovy, A.G., G.C. Angenent, H.J.M. Dons and A.C. van Altvorst, 1999. Heterologous expression of the *Arabidopsis etr1-1* allele inhibits the senescence of carnation flowers. Mol. Breed., 5: 301-308.
- 37. Gubrium, E.K., D.J. Clevenger, D.G. Clark, J.E. Barrett and T.A. Nell, 2000. Reproduction and horticultural performance of transgenic ethylene-insensitive petunias. J. Am. Soc. Hortic. Sci., 125: 277-281.
- 38. Ma, Y.P., L. Zhao, W.J. Zhang, Y.H. Zhang and X. Xing *et al.*, 2020. Origins of cultivars of *Chrysanthemum*-evidence from the chloroplast genome and nuclear *LFY* gene. J. Syst. Evol., 58: 925-944.
- 39. Inokuma, T., T. Kinouchi and S. Satoh, 2008. Reduced ethylene production in transgenic carnations transformed with ACC oxidase cDNA in sense orientation. J. Appl. Hortic., 10: 3-7.
- 40. Raffeiner, B., M. Serek and T. Winkelmann, 2009. *Agrobacterium tumefaciens*-mediated transformation of *Oncidium* and *Odontoglossum* orchid species with the ethylene receptor mutant gene *etr1-1*. Plant Cell, Tissue Organ Culture, 98: 125-134.
- 41. Sasaki, K., K. Kato, H. Mishima, M. Furuichi and I. Waga *et al.*, 2014. Generation of fluorescent flowers exhibiting strong fluorescence by combination of fluorescent protein from marine plankton and recent genetic tools in *Torenia fournieri* Lind. Plant Biotechnol., 31: 309-318.