



# Monoterpenoids and Their Synthesized Brominate Derivatives as Eco-Friendly Measures to Control Some Plant Pathogenic Fungi and Bacteria

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# ABSTRACT

Background and Objective: Phyto-pathogenic fungi or bacteria are one of the most important causes that decrease food and cash crops. Chemical pesticides cause damage to the environment. Thus, globally, there are attempts to provide eco-friendly products. Therefore, this work aimed to find alternative natural products for managing serious diseases (fungi and bacteria). Materials and Methods: Fungicidal and bactericidal of four monoterpenoids, as well as their synthesized brominated (Br) derivatives were investigated against six fungi and four bacteria in vitro. The synthesized brominated derivatives were confirmed by a mass spectrometer and GC/MS. Results: The soil and air-borne fungi were very sensitive to chlorothymol followed by thymol and carvacrol, respectively. However, the synthesized brominate derivatives such as bromocarvacrol were the most effective treatment against all the tested fungal strains, while, bromoegunol was a less effective treatment. Concerning the antibacterial activity, the tested monoterpenoids exhibited variable degrees of antibacterial against all of the tested bacterial strains and the conversion of monoterpenoids into bromo-derivatives enhanced the bactericidal activity. Conclusion: The chlorothymol (monoterpene) recorded the highest fungicidal activity against soil and airborn fungi. For bacterial activity, results exhibited that chlorothymol was the most effective monoterpene against bacteria. All the bromo-derivatives showed high activity against the most of tested fungi and bacteria.

## **KEYWORDS**

Fungicidal activity, bactericidal activity, monoterpenoids, brominate derivatives, soil-borne diseases, air-borne diseases

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# INTRODUCTION

Biopesticides are considered a unique solution to the problem of agrochemicals in crop protection. It reaches the target pest leaving the remaining 99.9% to enter the environment to cause hazards to non-target organisms including humans<sup>1</sup>. In the last two decades, scientists all over the world tried to minimize



the use of synthetic pesticides for the management of plant pathogens, insects, acari and weeds to avoid environmental pollution hazards. Besides, the targeted pathogen, pesticides may also kill various beneficial organisms. The increasing incidence of resistance among pathogens towards synthetic chemicals is also a cause of serious concern<sup>2</sup>. Major food and cash crops are attacked by certain pathogens especially fungal diseases which cause approximately 20% reductions in the yield<sup>3,4</sup>.

The discovery of antimicrobial compounds from plant sources is a safe route to a new generation characterized by eco-friendly, low toxicity, selectivity, highest efficacy and biodegradability<sup>5-7</sup>.

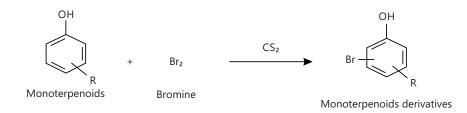
Search extensively for new biologically active terpenoids as a potential source for agrochemicals. Monoterpenes together with sesquiterpenes and diterpenes form the majority of essential oil. Monoterpenes are unsaturated hydrocarbons and some are oxygenated derivatives such as alcohol, ketones, carboxylic acid and phenol<sup>8,9</sup>. Chemical modification of natural monoterpenoids to various derivatives has been reported to result in modification of biological activity<sup>10-14</sup>. Therefore, this work was one of several attempts that have been conducted to find alternative natural products which are safe, biodegradable and eco-friendly for managing serious pests (fungi and bacteria) which attack crop plants.

## **MATERIALS AND METHODS**

In this investigation, the evaluated monoterpenoids and/or their synthesized brominated derivatives were implemented under laboratory conditions in 2021 at the Fungicide Bioassay Laboratory, Department of Pesticide Chemistry and Technology, Faculty of Agriculture, University of Alexandria, Alexandria, Egypt.

**Tested monoterpenoids:** The tested monoterpenoids were thymol, chlorothymol, eugenol and carvacrol. All monoterpenoids were purchased from Aldrich Chemicals Ltd., UK.

**General procedure for bromination of tested monoterpenoids:** The desired monoterpenoids (0.01 mol) thymol, eugenol, chlorothymol and carvacrol were dissolved in carbon disulfide (25 mL) in a round bottom flask fitted with a reflux condenser and pressure equivalent-dropping funnel. Bromine (0.01 mol) was dissolved in carbon disulfide (16 mL) and then added dropwise to a cooled flask (below 5°C) after the reaction mixture was stirred at room temperature for 1hr to complete (Scheme 1) and then monitored by thin layer chromatography (TLC with aluminum plate of 25, silica gel coated with fluorescent indicator F254, TLC size is 20×20 cm, MERCK, Germany). The reaction was quenched with water and extracted three times with methylene chloride (30 mL each). The organic layer was washed three times with water (100 mL) and dried over anhydrous sodium sulfate (50 g). The organic solvent was removed using a rotary evaporator under a high vacuum (brand name: Kryqcn, Model: RE501, rotating speed: 120 rpm, vacuum power: 0.098 mpa, China). The product was identified by a mass spectrometer (HP model, MS-5988) and GC/MS (Shimadzu QP2010) and Melting points were measured with capillary melting point apparatus model 1002-USA, UV spectra were recorded on a UV Spectrophotometer model UV-1601 SHIMADZU shown in Table 1 and 2.



Scheme 1: Synthesis of the monoterpenoidal brominate derivatives

Table 1: Physical properties of the monoterpenoidal derivatives

Derivative	Purity (%)	M.P (°C )
Bromothymol	87	48
Bromochlorothymol	85	Oil
Bromocarvacrol	97	46
Bromoeugenol	95	Oil

Compound		Empirical formula	Estimated M.W.	Found M.W.
Bromothymol	Br	C <sub>10</sub> H <sub>13</sub> OBr	229.11	229
Bromocholrothymol	CI Br OR	$C_{10}H_{12}OclBr$	263.56	263
Bromocarvacrol	Br	$C_{10}H_{13}Obr$	229.11	229
Bromoeugenol	Br OH	$C_{10}H_{11}O_2Br$	243.10	243

M.W.: Molecular weight

**Antifungal assay:** Six economic plant pathogenic fungi were chosen for this study. Fungi were obtained from the Fungicide Bioassay Laboratory, Department of Pesticide Chemistry, Faculty of Agriculture, Alexandria University. However, the studied fungi were as follows:

- Soil borne fungi: Fusarium oxyporum, Rhizoctonia solani and Pythium debrinum
- Airborne fungi: Alternaria alternata, Helminthosporium sp. and Botrytis feba

The antifungal activity of tested compounds was investigated by using the radial growth technique method<sup>15</sup>. Appropriate volumes of the stock solutions of the tested compounds either in dimethyl sulfoxide (DMSO) for solvent extracts or in distilled water for water extracts were added to molten nutrient agar (potato dextrose agar medium, PDA) to achieve the desired concentration immediately before pouring into the Petri dishes (9.0 cm in diameter) at 40-45°C.

Each concentration was tested in triplicate. Parallel controls were maintained with DMSO mixed with PDA. The discs of mycelial felt (0.5 cm diameter) of the plant pathogenic fungi, taken from 8-day-old cultures on PDA plates, were transferred aseptically to the center of Petri dishes. The treatments were incubated at 25°C in the dark. Colony growth diameter was measured after the fungal growth in the control treatments which completely covered the Petri dishes. Moreover, the percentage of mycelial growth inhibition was calculated according to the following formula<sup>16</sup>:

Mycelial growth inhibition = 
$$\frac{DC - DT}{DC} \times 100$$

where, DC and DT are the average diameters of the fungal colony of control and treatment, respectively.

**Bactericidal activity assay:** Four phytopathogenic bacteria, *Agrobacterium tumefaciens* (Crown gall bacteria), *Erwinia carotovra* subsp. *Carotovra* (soft rot bacteria), *Erwinia amylovora* (fire blight bacteria) and *Pseudomonas solanacearum* were provided by the Department of Plant Pathology, Faculty of Agriculture, Alexandria University, Egypt. The bacterial strains were cultured in a glycerol agar medium. Streptomycin sulfate as a standard bactericide was supplied by El-Nile Company for Chemical Industry and Drugs, Egypt.

**Determination of minimum inhibitory concentrations (MICs):** Tested compounds were dissolved in dimethyl sulfoxide (DMSO). Appropriate volumes of the stock solution were added to nutrient agar and poured into Petri dishes (diameter 9 mm). After solidification, a bacterial culture has grown in a nutrient broth for 18 hrs (approximately 10<sup>8</sup> CFU mL<sup>-1</sup> was planted on four lines per each plate with three replicates from four bacteria species *Agrobacterium tumefaciens, Erwinia carotovra, Erwinia amylovora* and *Pseudomonas salnacearum* on the surface of the agar. The inoculum line was allowed to dry before inverting the plates for incubation at 27°C for 24 hrs. The control was nutrient agar with a maximum volume of dimethylsulfoxide added to the treatment. The antibiotic, streptomycin was used for comparison, The MIC was determined as the lowest concentration of the tested compound showing no visible bacterial growth in the agar plates. As recommended by the European Society of Clinical Microbiology and Infection Disease<sup>17</sup>.

**Statistical analysis:** The effective concentrations of evaluated compounds that inhibit the fungi mycelia or bacteria growth by 50% ( $EC_{50}$ ), were conducted by a linear regression method with probit analysis<sup>18</sup>.

#### **RESULTS AND DISCUSSION**

**Identification of mono terpenoids derivatives:** The reaction of thymol, carvacrol, chlorothymol and eugenol with carbon disulfide and bromine gave bromothymol and bromocarvacrol as crystals, while bromocholorothymol and bromoeugenol as oily compounds. The percentage of the product yield obtained from the reaction was 80, 83, 92 and 90% for Bromothymol, Bromochlorothymol, Bromocarvacrol and Bromoeugenol, respectively. Physical properties and identification of the synthesized monoterpenoidal brominate derivatives showed in Table (1, 2) and Fig. (1-4).

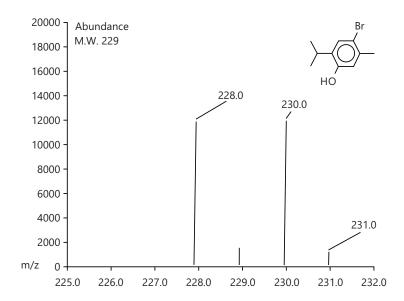


Fig. 1: MS spectra of bromothymol

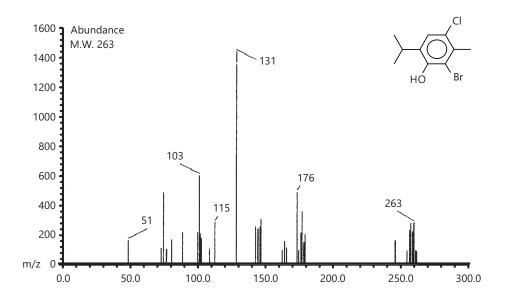


Fig. 2: MS spectra of bromocholorothymol

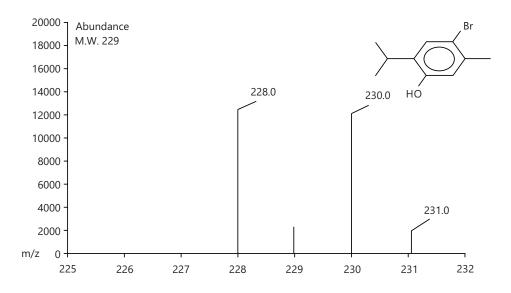
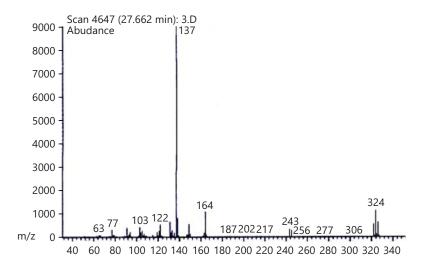


Fig. 3: MS spectra of bromocarvacrol

**Fungicidal activity of some monoterpenoids towards soil and air-borne fungi:** The fungicidal activity of four monoterpenoids namely, carvacrol, chlorothymol, eugenol and thymol were tested against soilborn fungi (*fusarium oxysprum*, *Rhizoctonia solani* and *Pythium debrinum*) and air-born fungi (*Botrytis feba*, *Helimanthosporum* sp. and *Alteranria alternate*) as shown in Table 3. The fungus *F. oxysproum* was very sensitive to chlorothymol, thymol, carvacrol and eugenol with EC<sub>50</sub> values estimated by 13.40, 40.57, 140.34 and 271.73 mg L<sup>-1</sup>, respectively, while *Rhizoctonia solani* was sensitive to chlorothymol (EC<sub>50</sub> = 21.00 mg L<sup>-1</sup>), carvacrol (EC<sub>50</sub> = 21.88 mg L<sup>-1</sup>), thymol (EC<sub>50</sub> = 30.90 mg L<sup>-1</sup>) and eugenol (EC<sub>50</sub> = 140.33). The soil-born fungus, *P. debrinum* recorded EC<sub>50</sub> values estimatedat 28.39, 43.00, 100.88 and 300 mg L<sup>-1</sup> with chlorothymol, thymol, carvacrol and eugenol, respectively. However, the standard fungicide (mancozeb) recorded EC<sub>50</sub> values of 140.34, 154.59 and 91.70 mg L<sup>-1</sup> with *F. oxysproum*, *R. solani* and *P. debrinum*, respectively.



# Fig. 4: MS spectra of bromoeugenol

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		95% confi	95% confidence limits	
Fungus	EC₅₀/ppm	Lower	Upper	Slope±SE
Chlorothymol				
Fusarium oxysporum	13.40	11.20	18.16	2.01±0.3
Rhizoctonia solani	21.00	15.50	31.22	2.00±0.19
Pythium debrinum	28.39	23.80	32.47	3.10±0.35
Alteranria altranata	07.58	05.31	8.44	1.10±0.18
Helimanthosporum sp.	04.49	00.21	7.09	2.60±0.35
Botrytis febae	03.80	00.84	5.12	1.40±0.16
Thymol				
Fusarium oxysporum	40.57	15.53	86.12	2.00±0.18
Rhizoctonia solani	30.90	27.14	34.84	2.60±0.24
Pythium debrinum	43.00	27.70	73.73	1.72±0.11
Alteranria altranata	27.82	15.61	42.70	2.80±0.26
Helimanthosporum sp.	17.38	09.66	100.14	2.30±0.29
Botrytis febae	22.97	08.85	40.88	2.40±0.3
Carvacrol				
Fusarium oxysporum	140.34	122.59	160.04	2.00±0.24
Rhizoctonia solani	21.85	16.59	27.44	1.60±0.19
Pythium debrinum	100.88	91.16	112.00	3.50±0.3
Alteranria altranata	18.80	12.40	25.80	0.78±0.13
Helimanthosporum sp.	53.96	22.92	93.62	1.90±0.2
Botrytis febae	23.99	14.98	30.97	1.60±32
Eugenol				
Fusarium oxysporum	271.73	217.01	403.75	2.68±0.4
Rhizoctonia solani	140.33	121.92	167.21	2.59±0.33
Pythium debrinum	>300	-	-	-
Alteranria altranata	>300	-	-	0.80±0.20
Helimanthosporum sp.	142.90	109.81	224.94	1.27±0.3
Botrytis febae	138.70	123.70	158.55	3.30±0.37
Mancozeb				
Fusarium oxysporum	140.34	122.59	160.04	2.00±0.24
Rhizoctonia solani	154.59	124.82	192.62	1.18±0.11
Pythium debrinum	91.70	61.93	294.80	0.90±0.28
Alteranria altranata	115.61	77.65	170.97	0.62±0.09
Helimanthosporum sp.	47.44	30.83	56.37	0.77±0.09
Botrytis febae	236.37	127.53	612.23	1.35±0.1

SE: Standard error

On the other hand, results showed that the air-born fungus *A. alternate* was very sensitive to chlorothymol  $(EC_{50} = 7.58 \text{ mg L}^{-1})$  followed by carvacrol  $(EC_{50} = 18.80 \text{ mg L}^{-1})$ , thymol  $(EC_{50} = 27.82 \text{ mg L}^{-1})$  and eugenol  $(EC_{50} = 300 \text{ mg L}^{-1})$ . *Helimanthosporum* sp., was sensitive towards chlorothymol, thymol, carvacrol and eugenol and recorded  $EC_{50}$  values estimated by 4.49, 17.38, 53.96 and 142.90 mg L<sup>-1</sup>, respectively. The airborn fungus *B. febae* was respectively very sensitive to chlorothymol, thymol, carvacrol and eugenol, with  $EC_{50}$  values of 3.80, 22.97, 23.99 and 138.70 mg L<sup>-1</sup>. The estimated  $EC_{50}$  values of mancozeb against *A. alternate, Helimanthosporum* sp. and *B. febae* were 115.61, 47.44 and 236.37 mg L<sup>-1</sup>, respectively.

The obtained results indicated that carvacrol, thymol, cholorothymol and eugenol exhibited the highest antifungal activity against phytopathogenic fungi<sup>19-22</sup>. The activity of thymol, carvacrol, chorothymol and eugenol against fungi may be attributed to containing the OH group in the aromatic ring. Also, the chlorine atom on the chlorothymol is playing an important role in the fungicidal activity against the tested fungal strains<sup>20,23</sup>. Therefore, it's possible to use these monoterpenoids as lead compounds to obtain new friendly fungicides for both humans and the environment.

**Fungicidal activity of synthesized monoterpenoidal brominate derivatives against soil and air-born fungi:** The fungicidal activity of brominate monoterpenoid derivatives against soil-born fungi i.e., *F. oxysporum, R. solani* and *P. debrinum*was shown in Table (4). The gained results indicated that *F. oxysporum* was sensitive to treatments of bromocarvacrol ( $EC_{50} = 21.86 \text{ mg L}^{-1}$ ), followed by Bromothymol ( $EC_{50} = 26.54 \text{ mg L}^{-1}$ ), bromoegunol ( $EC_{50} = 56.22 \text{ mg L}^{-1}$ ) and bromochlorothymol ( $EC_{50} = 60.10 \text{ mg L}^{-1}$ ). While *Rhizoctonia solani* showed high sensitivity towards bromochlorothymol, bromocarvacrol, bromothymol and bromoegunol with  $EC_{50}$  values of 9.38, 10.46, 28.08 and 40.51 mg L<sup>-1</sup>, consecutively. The fungus *P. debrinum* was very sensitive to the treatments of bromocarvacrol, bromochlorothymol, bromothymol and bromoegunol with  $EC_{50}$  values of 27.50, 40.93, 47.70 and 165.20 mg L<sup>-1</sup>, respectively. The standard fungicide (mancozeb) recorded  $EC_{50}$  values of 12.02, 91.70 and 154.60 mg L<sup>-1</sup> with *F. oxysporum, R. solani* and *P. debrinum*, respectively.

The air-born fungi results clarified that *A. alternata* was very sensitive to bromocarvacrol, bromothymol and bromochlorothymol with  $EC_{50}$  values of 16.34, 21.88 and 21.94 mg L<sup>-1</sup>, respectively, while it was less sensitive to bromoegunol ( $EC_{50}$  = 163.69 mg L<sup>-1</sup>). In the same context, *Helminthsporum* sp., recorded  $EC_{50}$  values estimated by 12.00, 15.47, 23.30 and 28.82 mg L<sup>-1</sup> with bromocarvacrol, bromochlorothymol, bromothymol and bromoegunol, respectively. *Botrytis fabae* showed sensitivity towards bromothymol ( $EC_{50}$  = 14.80 mg L<sup>-1</sup>), bromocarvacrol ( $EC_{50}$  = 15.36 mg L<sup>-1</sup>), bromochlorothymol ( $EC_{50}$  = 28.11 mg L<sup>-1</sup>) and bromoegunol ( $EC_{50}$  = 39.04 mg L<sup>-1</sup>). The treatment of mancozeb recorded  $EC_{50}$  values of 47.44, 115.61 and 236.37 mg L<sup>-1</sup> with *Helminthsporum* sp., *A. alternata* and *B. fabae*, respectively. These findings are in agreement with those obtained by Kaur *et al.*<sup>14</sup>, who found that halogenated thymol derivatives e.g. chlorothymol, dichlorothymo, monobromothymol and dibromothymol have strong antifungal and antibacterial activities. Also, Chauhan *et al.*<sup>24</sup>, found that thymol and its derivatives succeeded to inhibit the mycelia growth of *Rhizoctonia solani* during the *in vitro* study, while the soil application under greenhouse conditions thymol or its derivatives suppressed the damping-off in cucumber seedlings at a range of 26.67 to 100%.

**Bactericidal activity of tested monoterpenoids and their brominate derivatives:** In the current investigation, the bactericidal activity of four monoterpenoid compounds namely, carvacrol, thymol, chlorothymol and eugenol, in addition to, their brominated derivatives, Bromochlorothymol, Bromothymol, Bromocarvacrol and Bromoeugenol were assessed against *A. tumefaciens*, *E. cartovora*, *E. amylovora* and *P. solanacearum* under laboratory conditions (Table 5). The bacterium of *A. tumefacient* was very sensitive to standard bactericide streptomycin with MIC = 5 µg mL<sup>-1</sup>, followed by *E. cartovora*, *E. amylovora* and *P. solanacearum* with values estimated by 10 µg mL<sup>-1</sup> of each.

			dence limits	
Fungus	EC <sub>50</sub> /ppm	Lower	Upper	Slope±SE
Bromochlorothymol				
Fusarium oxysprum	60.10	49.90	72.10	1.21±0.11
Rhizoctonia solani	9.38	7.80	10.90	2.30±0.24
Pythium debrinum	40.93	33.40	48.60	1.97±0.30
Alteranria altranata	21.94	1.25	40.30	2.00±0.24
Helimanthosporum sp.	15.47	9.40	46.80	2.50±0.29
Botrytis febae	28.11	15.77	50.11	1.00±0.10
Bromothymol				
Fusarium oxysprum	26.54	18.57	39.73	3.00±0.23
Rhizoctonia solani	28.08	11.54	150.56	2.30±0.27
Pythium debrinum	47.70	31.07	69.17	1.30±0.30
Alteranria altranata	21.88	17.17	33.35	1.40±0.26
Helimanthosporum sp.	23.30	20.26	29.50	2.80±0.15
Botrytis febae	14.80	9.47	23.36	2.80±0.23
Bromocarvacrol				
Fusarium oxysprum	21.86	16.59	27.44	1.63±0.15
Rhizoctonia solani	10.46	9.14	12.23	2.55±0.4
Pythium debrinum	27.50	23.29	33.626	1.82±0.19
Alteranria altranata	16.34	14.42	18.57	2.25±0.2
Helimanthosporum sp.	12.00	10.40	14.60	2.40±0.4
Botrytis febae	15.36	12.64	18.71	4.00±0.31
Bromoeugenol				
Fusarium oxysprum	56.22	28.70	70.20	2.70±0.25
Rhizoctonia solani	40.51	18.78	61.32	2.00±0.19
Pythium debrinum	165.2	139.00	200.2	1.70±0.23
Alteranria altranata	163.69	138.32	198.30	1.70±0.23
Helimanthosporum sp.	28.82	11.81	51.42	1.14±0.11
Botrytis febae	39.04	20.88	61.65	1.30±0.12
Mancozeb				
Fusarium oxysprum	12.02	4.59	25.82	1.38±0.06
Rhizoctonia solani	154.60	124.82	192.62	1.18±0.11
Pythium debrinum	91.70	61.93	294.80	0.90±0.20
Alteranria altranata	115.61	77.65	170.97	0.62±0.09
Helimanthosporum sp.	47.44	30.83	56.37	0.77±0.09
Botrytis febae	236.37	127.53	612.23	1.35±0.1

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Lable 4. Filbolcidal activit	y of tested monoterpenoic	i derivatives adains	t nnvtonathogenic tungi

SE: Standard error

Table 5: Bactericidal activity of some monoterpenoids and their derivatives by minimal inhibitory concentration (MIC)

Compound (μg mL <sup>-1</sup> )	A. tumefaciens	E. amylovora	E. cartovora	P. solanacearm
Chlorothymol	35	50	35	60
Thymol	100	90	300	>300
Eugenol	220	300	>300	300
Carvacrol	45	50	50	50
Bromochlorothymol	5	50	5	60
Bromothymol	9	200	10	3
Bromoeugenol	5	50	5	5
Bromocarvacrol	5	5	5	5
Sterptomycin	5	10	10	10

With respect to the *A. tumefaciens* strain it was sensitive to chlorothymol, carvacrol, thymol and eugenol with values of 35, 45, 100 and 220  $\mu$ g mL<sup>-1</sup>, respectively. Meanwhile, *A. tumefaciens* showed great sensitivity towards bromochlorothymol, bromocarvacrol, bromoeugenol and bromothymol with MIC values of 5, 5, 5 and 9  $\mu$ g mL<sup>-1</sup>, respectively. However, *E. amylovora* strain exhibited sensitivity to chlorothymol, carvacrol, thymol and eugenol with MIC values of 50, 50, 50 and 300  $\mu$ g mL<sup>-1</sup>, respectively. Whilst, *E. amylovora* recorded sensitivity towards bromocarvacrol (5  $\mu$ g mL<sup>-1</sup>), bromochlorothymol (50  $\mu$ g mL<sup>-1</sup>) and bromothymol (200  $\mu$ g mL<sup>-1</sup>).

On the other hand, *E. cartovora* recorded sensitivity towards tested monoterpenoids such as chlorothymol, carvacrol, thymol and eugenol with values of 35, 50, 300 and >300  $\mu$ g mL<sup>-1</sup>, respectively. However, *E. cartovora* exhibited high sensitivity to bromochlorothymol (5  $\mu$ g mL<sup>-1</sup>), bromocarvacrol (5  $\mu$ g mL<sup>-1</sup>) and bromothymol (10  $\mu$ g mL<sup>-1</sup>). According to the obtained data, *P. solanacearum* showed sensitivity towards carvacrol, chlorothymol, eugenol and thymol with MIC values estimated by 50, 60, 300 and >300, consecutively. Furthermore, the transformed monoterpenoids such as bromothymol, bromocarvacrol, bromoeugenol and bromochlorothymol were very effective against *P. solanacearum* achieving 3, 5, 5 and 60  $\mu$ g mL<sup>-1</sup>, respectively.

Several studies reported that the monoterpenes exhibited antifungal and antibacterial activities against a wide range of microorganisms<sup>25-28</sup>. The results of the current investigation are in agreement with El-Zemity *et al.*<sup>29</sup> who stated that chlorothymol was effective as a bactericide against *Agrobacterium tumefaciens* and *Erwinia carotovora var carotovora*. The activity of chlorothymol may be attributed to the chlorine atom which plays a good role in the bactericidal activity against the tested bacteria. Also, Mahboub and Memmou<sup>30</sup> found that synthesized 6-bromoeugenol or eugenol has antibacterial activity, especially in both positive and negative gram bacteria. Also, thymol derivatives had antibacterial activities with minimum inhibitory concentration values, 40 to 80 µg mL<sup>-131</sup>. Thymyl-4-nitrobenzoate recorded good inhibitory action towards plant pathogenic bacteria either as a major constituent or in combination with other antimicrobial agents<sup>32</sup>. Abd-El-Aziz *et al.*<sup>33</sup> indicated that using the essential oils of caraway (*Carum carvi* L.) and thyme (*Thymus vulgaris* L.) exhibited antibacterial efficacy against *Agrobacterium tumefaciens* in a laboratory study. Moreover, the application of caraway and thyme oils suppressed the gall formation by 63.16 and 89.47%, respectively, on apricot seedlings in pots.

## CONCLUSION

From the above results, it could be concluded that chlorothymol (monoterpene) recorded the highest fungicidal activity against the most of soil and air-born fungi. While, the brominated derivatives of monoterpenoids showed that bromocarvacrol was the most effective against all tested fungi, whereas, bromoegunol was the least one. The same trend was recorded with bacteria, where chlorothymol (monoterpene) recorded the highest bactericidal activity. However, the brominated derivatives of monoterpenoids were more effective against all the tested bacteria. Therefore, this study proposed some eco-friendly solutions, but more experiments are needed in the future.

## SIGNIFICANCE STATEMENT

Fungi and bacteria are among the most common plant pathogens that lead to losses in agricultural crops, whether during planting or after harvest. Combating these diseases has become costly, especially recently, so it was necessary to search for effective alternatives that can be used safely without any adverse effects. This study was thus conducted to assess the impact of some monoterpenes and their brominate derivatives against soil and air fungi as well as bacteria. Also, results showed that some of the monoterpenes or their derivatives e.g., chlorothymol and bromocarvacrol were effective as fungicides and/or bactericides.

## REFERENCES

- Chandler, D., A.S. Bailey, G.M. Tatchell, G. Davidson, J. Greaves and W.P. Grant, 2011. The development, regulation and use of biopesticides for integrated pest management. Philos. Trans. R. Soc. B: Biol. Sci., 366: 1987-1998.
- 2. Heydari, A. and M. Pessarakli, 2010. A review on biological control of fungal plant pathogens using microbial antagonists. J. Biol. Sci., 10: 273-290.
- 3. Omezzine, F., M. Bouaziz, M. Daami-Remadi, M.S.J. Simmonds and R. Haouala, 2017. Chemical composition and antifungal activity of *Trigonella foenum-graecum* L. varied with plant ploidy level and developmental stage. Arabian J. Chem., 10: S3622-S3631.

- 4. Shuping, D.S.S. and J.N. Eloff, 2017. The use of plants to protect plants and food against fungal pathogens: A review. Afr J. Tradit. Complementary Altern. Med., 14: 120-127.
- Silvan, J.M., E. Guerrero-Hurtado, A. Gutierrez-Docio, M. Prodanov and A.J. Martinez-Rodriguez, 2022. Olive leaf as a source of antibacterial compounds active against antibiotic-resistant strains of *Campylobacter jejuni* and *Campylobacter coli*. Antibiotics, Vol. 12. 10.3390/antibiotics12010026.
- Chodkowska, K.A., H. Iwiński, K. Wódz, T. Nowak and H. Różański, 2022. *In vitro* assessment of antimicrobial activity of phytobiotics composition towards of avian pathogenic *Escherichia coli* (APEC) and other *E. coli* strains isolated from broiler chickens. Antibiotics, Vol. 11. 10.3390/antibiotics11121818.
- Vaou, N., E. Stavropoulou, C. Voidarou, C. Tsigalou and E. Bezirtzoglou, 2021. Towards advances in medicinal plant antimicrobial activity: A review study on challenges and future perspectives. Microorganisms, Vol. 9. 10.3390/microorganisms9102041.
- 8. Zielińska-Błajet, M. and J. Feder-Kubis, 2020. Monoterpenes and their derivatives-recent development in biological and medical applications. Int. J. Mol. Sci., Vol. 21. 10.3390/ijms21197078.
- Stephane, F.F.Y. and B.K.J. Jules, 2020. Terpenoids as Important Bioactive Constituents of Essential Oils. In: Essential Oils - Bioactive Compounds, New Perspectives and Applications, de Oliveira, M.S., S. Silva and W.A. da Costa (Eds.), IntechOpen, London, UK, ISBN: 978-1-83962-698-2, pp: 222.
- 10. Marinelli, L., A. di Stefano and I. Cacciatore, 2018. Carvacrol and its derivatives as antibacterial agents. Phytochem. Rev., 17: 903-921.
- 11. Wang, Y., J. Ren, X.H.H. Huang, R. Tong and J.Z. Yu, 2017. Synthesis of four monoterpene-derived organosulfates and their quantification in atmospheric aerosol samples. Environ. Sci. Technol., 51: 6791-6801.
- 12. Wang, K., S. Jiang, T. Pu, L. Fan, F. Su and M. Ye, 2019. Antifungal activity of phenolic monoterpenes and structure-related compounds against plant pathogenic fungi. Nat. Prod. Res., 33: 1423-1430.
- 13. Cui, Z., X. Li and Y. Nishida, 2014. Synthesis and bioactivity of novel carvacrol and thymol derivatives containing 5-phenyl-2-furan. Lett. Drug Des. Discovery, 11: 877-885.
- 14. Kaur, R., M.P. Darokar, S.K. Chattopadhyay, V. Krishna and A. Ahmad, 2014. Synthesis of halogenated derivatives of thymol and their antimicrobial activities. Med. Chem. Res., 23: 2212-2217.
- 15. Selim, R.E., S.M. Ahmed, S.R. El-Zemity, S.S. Ramses and Y.T.A. Moustafa, 2015. Antifungal activity and seasonal variation of green alga (*Ulva lactuca*) extracts. Asian J. Agric. Food Sci., 3: 419-427.
- 16. Tian, L., X. Chen, X. Jia, S. Wang and X. Wang *et al.*, 2021. First report of antifungal activity conferred by non-conventional peptides. Plant Biotechnol. J., 19: 2147-2149.
- 17. European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID), 2000. Determination of minimum inhibitory concentrations (MICs) of antibacterial agents by agar dilution. Clin. Microb. Infect., 6: 509-515.
- 18. Finney, D.J., 1971. Probit Analysis. 3rd Edn., Cambridge University Press, Cambridge, ISBN: 9780521080415, Pages: 333.
- 19. Numpaque, M.A., L.A. Oviedo, J.H. Gil, C.M. García and D.L. Durango, 2011. Thymol and carvacrol: Biotransformation and antifungal activity against the plant pathogenic fungi *Colletotrichum acutatum* and *Botryodiplodia theobromae*. Trop. Plant Pathol., 36: 3-13.
- 20. Kordali, S., A. Cakir, H. Ozer, R. Cakmakci, M. Kesdek and E. Mete, 2008. Antifungal, phytotoxic and insecticidal properties of essential oil isolated from Turkish *Origanum acutidens* and its three components, carvacrol, thymol and *p*-cymene. Bioresour. Technol., 99: 8788-8795.
- Lee, O.G., G.J. Choi, K.S. Jang, H.K. Lim, K.Y. Cho and J.C. Kim, 2007. Antifungal activity of five plant essential oils as fumigant against postharvest and soilborne plant pathogenic fungi. Plant Pathol. J., 23: 97-102.
- 22. Tsao, R. and T. Zhou, 2000. Antifungal activity of monoterpenoids against postharvest pathogens *Botrytis cinerea* and *Monilinia fructicola*. J. Essent. Oil Res., 12: 113-121.

- 23. Abbaszadeh, S., A. Sharifzadeh, H. Shokri, A.R. Khosravi and A. Abbaszadeh, 2014. Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. J. Mycol. Med., 24: e51-e56.
- 24. Chauhan, K.R., T.C. Le, P.K. Chintakunta and D.K. Lakshman, 2017. Phyto-fungicides: Structure activity relationships of the thymol derivatives against *Rhizoctonia solani*. J. Agric. Chem. Environ., 6: 175-185.
- 25. Marchese, A., I.E. Orhan, M. Daglia, R. Barbieri and A.D. Lorenzo *et al.*, 2016. Antibacterial and antifungal activities of thymol: A brief review of the literature. Food Chem., 210: 402-414.
- 26. de Castro Teixeira, A.P., R. de Oliveira Nóbrega, E. de Oliveira Lima, W. de Oliveira Araújo and I. de Oliveira Lima, 2020. Antifungal activity study of the monoterpene thymol against *Cryptococcus neoformans*. Nat. Prod. Res., 34: 2630-2633.
- 27. Zhou, L., Z. Zhang, M. Wei, Y. Xie, S. He, H. Shi and Z. Lin, 2019. Evaluation of the antifungal activity of individual and combined monoterpenes against *Rhizopus stolonifer* and *Absidia coerulea*. Environ. Sci. Pollut. Res., 26: 7804-7809.
- Ahmed, B., A. Jailani, J.H. Lee and J. Lee, 2022. Inhibition of growth, biofilm formation, virulence, and surface attachment of *Agrobacterium tumefaciens* by cinnamaldehyde derivatives. Front. Microbiol., Vol. 13. 10.3389/fmicb.2022.1001865.
- 29. El-Zemity, S.R., M.A. Radwan, S.A.E.M. Mohamed and S.M. Sherby, 2008. Antibacterial screening of some essential oils, monoterpenoids and novel *N*-methyl carbamates based on monoterpenoids against *Agrobacterium tumefaciens* and *Erwinia carotovora*. Arch. Phytopathol. Plant Protect., 41: 451-461.
- 30. Mahboub, R. and F. Memmou, 2016. Antimicrobial properties of 6-bromoeugenol and eugenol. Int. Lett. Nat. Sci., 53: 57-64.
- 31. Swain, S.S., S.K. Paidesetty and R.N. Padhy, 2019. Synthesis of novel thymol derivatives against MRSA and ESBL producing pathogenic bacteria. Nat. Prod. Res., 33: 3181-3189.
- 32. Kumar, A., S.P. Singh and S.S. Chhokar, 2008. Thymol and its derivatives as antimicrobial agents. Nat. Prod. Commun., Vol. 3. 10.1177/1934578X0800300528.
- 33. Abd-El-Aziz, R.M., A.F. Abd El-Rahman and D.M.G. Hendi, 2021. Control of *Agrobacterium tumefaciens* with essential oils compared to antagonistic *Agrobacterium radiobacter* strain K84. Egypt. J. Phytopathol., 49: 80-92.